

## Zinc ions block the second but not the first phasic response to repetitive application of carbachol and histamine in guinea-pig taenia caeci

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**Abstract.** In the presence of  $\text{Zn}^{2+}$  (0.3 mM), carbachol ( $10^{-6}$  M) or histamine ( $10^{-5}$  M) induced the phasic response in guinea-pig taenia caeci while the tonic response was markedly inhibited. However, when the muscles were kept in  $\text{Zn}^{2+}$ -containing medium following the first stimulation with either carbachol or histamine, neither application of carbachol nor of histamine elicited another phasic contraction. Caffeine (25 mM) did not induce contraction in the presence of  $\text{Zn}^{2+}$ . After the washing out of caffeine in the presence of  $\text{Zn}^{2+}$ , however, the muscle did then develop the phasic response on the application of carbachol or histamine. In conclusion,  $\text{Zn}^{2+}$  did not affect the carbachol or histamine-induced  $\text{Ca}^{2+}$  release from the storage sites. However, when  $\text{Zn}^{2+}$  was continuously present,  $\text{Ca}^{2+}$  was not supplied to the storage sites. Furthermore, carbachol and histamine mobilized a common cellular  $\text{Ca}^{2+}$  store, but they activated  $\text{Ca}^{2+}$  release channels different from the ones activated by caffeine in the  $\text{Ca}^{2+}$  storage sites.

**Key words.**  $\text{Zn}^{2+}$ ; calcium store; smooth muscle.

Zinc ions ( $\text{Zn}^{2+}$ ) have been reported to inhibit contractions due to acetylcholine, histamine and  $\text{K}^+$  in ileal longitudinal smooth muscle<sup>1-3</sup>. I have shown earlier<sup>3</sup> that  $\text{Zn}^{2+}$  did not affect the phasic response to  $\text{K}^+$  (60 mM), but it inhibited the  $\text{K}^+$  tonic tension mainly by inhibition of  $\text{Ca}^{2+}$  influx via the voltage-operated  $\text{Ca}^{2+}$  channels. However, when  $\text{Zn}^{2+}$  was present in the external surroundings,  $\text{Ca}^{2+}$  was not supplied to the storage sites and was not available for the second phasic response to  $\text{K}^+$ .

It is well known that vascular smooth muscle contractions in response to norepinephrine, histamine and caffeine depend on  $\text{Ca}^{2+}$  release from cellular  $\text{Ca}^{2+}$  storage sites<sup>4-7</sup>. Also in intestinal muscle, carbachol and caffeine induce  $\text{Ca}^{2+}$  release from  $\text{Ca}^{2+}$  storage sites<sup>8,9</sup>. It has been supposed that in the taenia caeci there exist two  $\text{Ca}^{2+}$  release mechanisms, namely  $\text{Ca}^{2+}$ -induced  $\text{Ca}^{2+}$  release (CICR), and inositol 1,4,5-triphosphate ( $\text{IP}_3$ )-induced  $\text{Ca}^{2+}$  release from the  $\text{Ca}^{2+}$  storage sites<sup>10,11</sup>.

The present study was undertaken to examine the effects of  $\text{Zn}^{2+}$  on phasic responses induced by  $\text{Ca}^{2+}$  release from  $\text{Ca}^{2+}$  stores by carbachol, histamine and caffeine in the taenia caeci.

### Materials and methods

Strips of the taenia caeci were isolated from the caecum of male Hartley strain guinea-pigs, b. wt 400 g, and were immersed in modified normal Tyrode solution bubbled with 100%  $\text{O}_2$  at 37 °C. The solution contained (mM): NaCl, 123.7; KCl, 2.7;  $\text{CaCl}_2$ , 2.5;  $\text{MgCl}_2$ , 1.0, tris(hydroxymethyl)aminomethane, 25; and glucose, 5.5. The pH of the solution was adjusted to 7.4 with

HCl at 37 °C. Zinc ions, as  $\text{ZnCl}_2$ , were directly added to the bath solution.

The muscle strips were suspended at a resting tension of 0.6 g and allowed to equilibrate for 40 min with several changes of the Tyrode solution. Isometric contraction of the muscle was measured by a strain gauge transducer (Nihon Kohden, RM-6000). After equilibration, the muscles were conditioned by adding 40 mM  $\text{K}^+$ .

### Results

When a strip of taenia caeci muscle was treated with carbachol ( $10^{-6}$  M) or histamine ( $10^{-5}$  M), each maximal phasic tension was followed by a sustained tonic contraction. The effects of preincubation with  $\text{Zn}^{2+}$  for 30 min on tension induced by high  $\text{K}^+$  (60 mM, hypertonic), carbachol and histamine were examined.  $\text{Zn}^{2+}$  (0.1 mM) had little effect on the phasic response to  $\text{K}^+$  and carbachol; however, the tonic responses to  $\text{K}^+$  and carbachol were reduced markedly, to  $38.1 \pm 4.3\%$  ( $n = 10$ ) and  $9.1 \pm 0.8\%$  ( $n = 10$ ) of the original tonic responses, respectively (fig. 1). In the presence of 0.1 mM  $\text{Zn}^{2+}$ , about 60% of the phasic response to histamine remained, but the tonic response slowly decayed completely to the baseline level. With a higher concentration of  $\text{Zn}^{2+}$  (0.3 mM),  $\text{K}^+$ , carbachol or histamine produced only a phasic response although the period was slightly shortened (fig. 2). However,  $\text{Zn}^{2+}$  blocked the second phasic response to repetitive application of carbachol or histamine (figs 1 and 2).

In the next series of experiments, after the first application of carbachol or histamine in the presence of  $\text{Zn}^{2+}$ , muscles were stimulated again with the other agonist. Carbachol (histamine) could not evoke a second con-

(A)

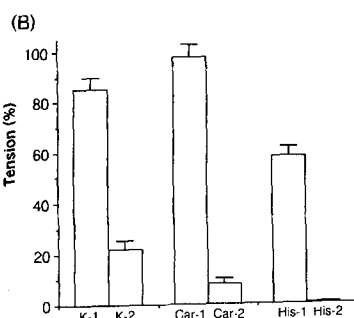
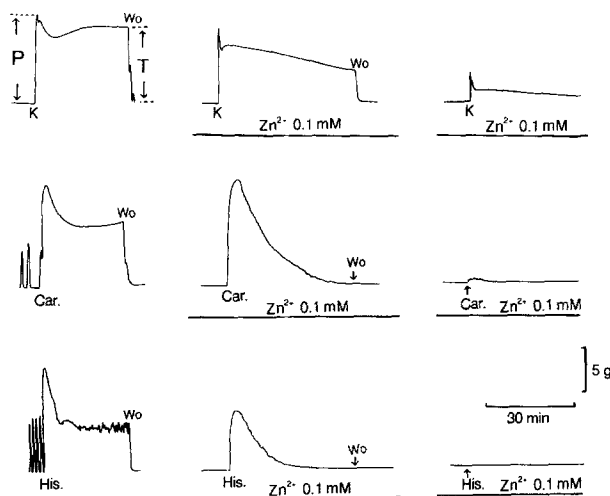


Figure 1. The effects of preincubation with  $Zn^{2+}$  on responses to  $K^+$ , carbachol and histamine in taenia coli.

A In each row, the first response to  $K^+$  (hypertonic, 60 mM), carbachol ( $1 \times 10^{-6}$  M) and histamine ( $1 \times 10^{-5}$  M) was the control. Muscles were preincubated for 30 min in 0.1 mM  $Zn^{2+}$  medium before application of  $K^+$ , carbachol and histamine. Following application of  $K^+$ , carbachol and histamine in the presence of 0.1 mM  $Zn^{2+}$  for 45 min, the muscles were washed with the medium containing 0.1 mM  $Zn^{2+}$ .  $K^+$ , carbachol and histamine were then reapplied in the presence of 0.1 mM  $Zn^{2+}$ .

K:  $K^+$  (60 mM); Car.: Carbachol ( $1 \times 10^{-6}$  M); His.: Histamine ( $1 \times 10^{-5}$  M); P: Phasic contraction; T: Tonic contraction; Wo: Wash-out.

B The first and second phasic responses stimulated by  $K^+$ , carbachol and histamine in the presence of 0.1 mM  $Zn^{2+}$  were calculated as percentages of phasic responses to  $K^+$ , carbachol and histamine in the absence of  $Zn^{2+}$ , respectively.

K-1, Car-1 or His-1: first phasic response to  $K^+$ , carbachol or histamine, respectively, in the presence of 0.1 mM  $Zn^{2+}$ . K-2, Car-2 or His-2: second phasic response to  $K^+$ , carbachol or histamine, respectively, in the presence of 0.1 mM  $Zn^{2+}$ . Error bars on the columns show SE of 10–12 experiments.

traction after the washing out of histamine (carbachol) in the presence of 0.3 mM  $Zn^{2+}$ , and vice versa (figs 3 and 4).

Caffeine (25 mM) induced a rapid transient contraction in taenia caeci strips (fig. 5). Caffeine has been shown to evoke a transient contraction by  $Ca^{2+}$  release from the store in the ileum<sup>8,9</sup>. In the presence of 0.1 mM  $Zn^{2+}$ , caffeine did not evoke any contraction. However, when  $K^+$ , carbachol or histamine was applied after the wash-

ing out of caffeine in the presence of 0.1 mM  $Zn^{2+}$ , the agonists produced phasic responses (fig. 5).

## Discussion

$Zn^{2+}$  markedly inhibited both the tonic response, and the  $Ca^{2+}$  binding at low affinity sites, induced by  $K^+$  (60 mM). There were smaller effects on the phasic response and the high affinity  $Ca^{2+}$  site in the taenia caeci<sup>3</sup>. It is well established that in taenia caeci the phasic response to  $K^+$  (40–60 mM) can be attributed to release of  $Ca^{2+}$  from a cellular store of  $Ca^{2+}$  binding

(A)

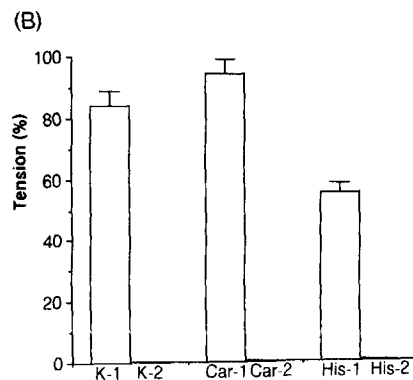
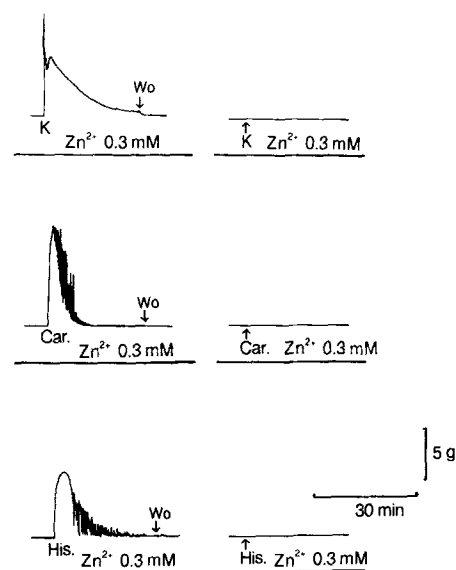


Figure 2. The effects of 0.3 mM  $Zn^{2+}$  on responses to  $K^+$  (60 mM), carbachol ( $10^{-6}$  M) and histamine ( $10^{-5}$  M).

A In each experiment muscle was treated in the same way as in figure 1A except that the concentration of  $Zn^{2+}$  was 0.3 mM. The control response to  $K^+$ , or carbachol and histamine in the absence of  $Zn^{2+}$  was omitted in figure.

B Responses were expressed as a percentage of the corresponding phasic response before addition of  $Zn^{2+}$ .

K-1, Car-1 or His-1: first phasic response to  $K^+$ , carbachol or histamine in the presence of 0.3 mM  $Zn^{2+}$ , respectively.  $Zn^{2+}$  (0.3 mM) completely blocked the second phasic responses (K-2, Car-2 or His-2) to  $K^+$ , carbachol or histamine, respectively, of the same agonist as first stimulation. Error bars on the columns show SE of 10–12 experiments.

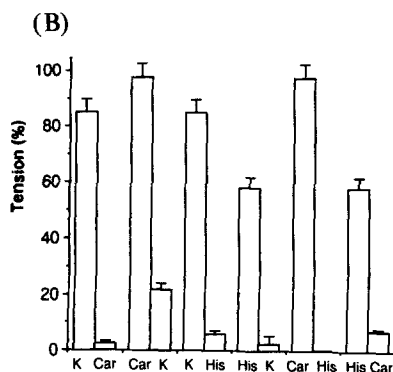
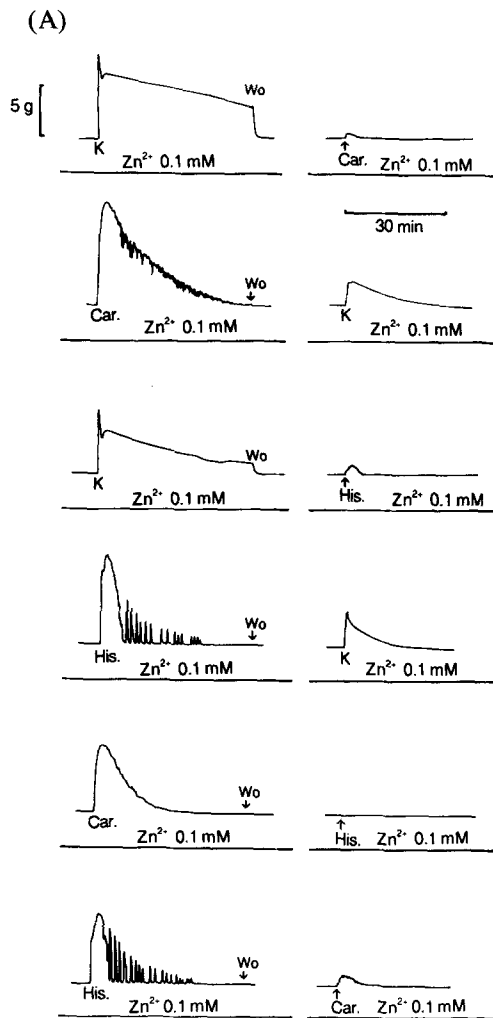


Figure 3. Effects of 0.1 mM  $Zn^{2+}$  on responses to second stimulation with a different agonist. The protocol and the concentration of each agonist of the experiments in A was the same as in figure 1A, respectively. The left bar in each row of graph B represents first phasic response to one agonist, the right bar is the second phasic response to another. Error bars on the columns show SE of 10 experiments.

sites at a high affinity, as was determined by the lanthanum method, and the tonic response can be attributed to increased inward movement of  $Ca^{2+}$  from extracellular fluid<sup>12-14</sup>. It is also known that carbachol and histamine activate muscarinic receptors and his-

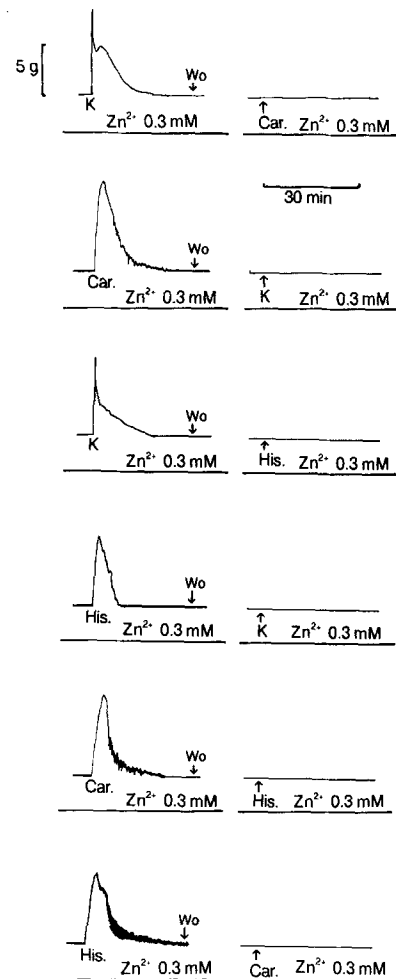


Figure 4. Effects of 0.3 mM  $Zn^{2+}$  on responses to second stimulation with a different agonist. The protocol and the concentration of each agonist of the experiments was the same as in figure 1A.

tamine receptors, respectively, and these receptor activations evoke a production of  $IP_3$  due to activity of triphosphoinositide phosphodiesterase<sup>15</sup>.  $IP_3$  induces  $Ca^{2+}$  release from the storage sites<sup>10,11,15,16</sup>, which results in an initial phasic contraction. This process is accompanied by the opening of receptor-linked  $Ca^{2+}$  channel in response to the acceleration of electrical membrane activity<sup>8,17</sup>, which results in a sustained tonic contraction. The selectivity for the inhibition of the tonic response by  $Zn^{2+}$  probably indicates that  $Zn^{2+}$  inhibited the receptor-linked  $Ca^{2+}$  channel but it did not affect the  $Ca^{2+}$  release from the storage sites activated by histamine and carbachol.

However,  $Zn^{2+}$  completely inhibited the second phasic response to repetitive and sequential application of carbachol and histamine. Both the phasic response and the high affinity  $Ca^{2+}$  sites induced by  $K^+$  in the presence of  $Zn^{2+}$  were non-responsive to a second stimulation with  $K^+$  in taenia caeci<sup>3</sup>. This probably indicates that these agonists (carbachol and histamine) depleted  $Ca^{2+}$  in the storage sites sensitive to them by the first stimulation,

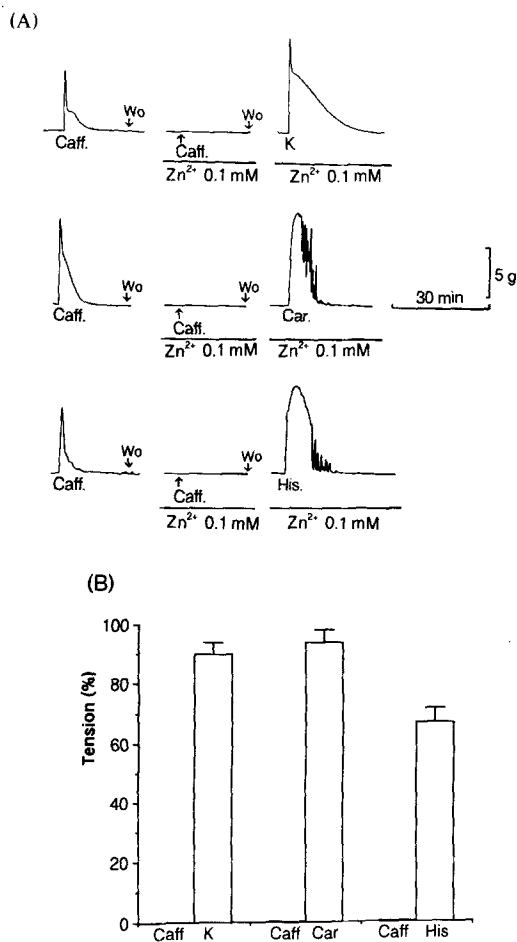


Figure 5. Effects of  $Zn^{2+}$  on response to caffeine.

A The incubation time of caffeine was 20 min. Caffeine (25 mM) did not induce the response in the presence of 0.1 mM  $Zn^{2+}$ , but after the caffeine had been washed away, the muscle developed first phasic responses to  $K^+$  (60 mM), carbachol ( $10^{-6}$  M) or histamine ( $10^{-5}$  M).

B The phasic response stimulated by  $K^+$ , carbachol and histamine after the washing out of caffeine in the presence of 0.1 mM  $Zn^{2+}$  were calculated as percentages of the phasic response to  $K^+$ , carbachol and histamine in the absence of  $Zn^{2+}$ . Error bars on the columns show SE of 10 experiments.

and  $Ca^{2+}$  was not refilled into the  $Ca^{2+}$  storage sites in the presence of  $Zn^{2+}$ .

Caffeine did not induce a contraction in the presence of a low concentration of  $Zn^{2+}$  (0.1 mM). It is probable that the inhibition of the caffeine response by  $Zn^{2+}$  is caused by inhibition of the ' $Ca^{2+}$ -induced  $Ca^{2+}$  release (CICR)'. However, when  $K^+$ , carbachol or histamine was applied after a wash with caffeine in the presence of 0.1 mM  $Zn^{2+}$ , the phasic response to the agonists could be produced (fig. 5). This may be because subsequent application of an agonist after caffeine application in the presence of  $Zn^{2+}$  produces  $IP_3$  and this  $IP_3$  releases  $Ca^{2+}$  via the  $IP_3$  channel instead of the CICR channel. In conclusion, it was suggested that  $Zn^{2+}$  did not affect carbachol or histamine-induced  $Ca^{2+}$  release from the store sites. However,  $Ca^{2+}$  was not supplied to the store sites in the presence of  $Zn^{2+}$  in ileal muscle. I think that these results suggest the utility of  $Zn^{2+}$  as a biochemical and pharmacological tool for testing the function of the intracellular  $Ca^{2+}$  store mobilized by agonists in intact smooth muscle cells.

- Schnieden, H., and Small, R. C., Br. J. Pharmac. 41 (1971) 488.
- Sarria, B., Cortijo, J., Marti-Cabrera, M., Morcillo, E., and Esplugues, J., Br. J. Pharmac. 97 (1989) 19.
- Nasu, T., Experientia 49 (1993) 865.
- Deth, R. C., and Lynch, C. J., Am. J. Physiol. 240 (1981) C239.
- Leijten, P. A. A., and Van Breemen, C., J. Physiol. 357 (1984) 327.
- Matsumoto, T., Kanaide, H., Shogakiuchi, Y., and Nakamura, M., J. Biol. Chem. 265 (1990) 5610.
- Noguera, M. A., and D'Ocon, M. P., Naunyn Schmiedeberg's Archs Pharmacol. 345 (1992) 333.
- Bolton, T. B., and Lim, S. P., J. Physiol. 409 (1989) 385.
- Pacaud, P., and Bolton, T. B., J. Physiol. 441 (1991) 477.
- Iino, M., Biochem. biophys. Res. Commun. 142 (1987) 47.
- Komori, S., and Bolton, T. B., J. Physiol. 433 (1991) 495.
- Urakawa, N., and Holland, W. C., Am. J. Physiol. 207 (1964) 873.
- Karaki, H., Nakagawa, H., and Urakawa, N., Br. J. Pharmac. 81 (1984) 393.
- Nasu, T., and Ishida, Y., Gen. Pharmac. 21 (1990) 349.
- Somlyo, A. V., Bond, M., Somlyo, A. P., and Scarpa, A., Proc. natl Acad. Sci. USA 82 (1985) 5231.
- Kobayashi, S., Gong, M. C., Somlyo, A. V., and Somlyo, A. P., Am. J. Physiol. 260 (1991) C364.
- Mitsui, M., and Karaki, H., Am. J. Physiol. 258 (1990) C787.